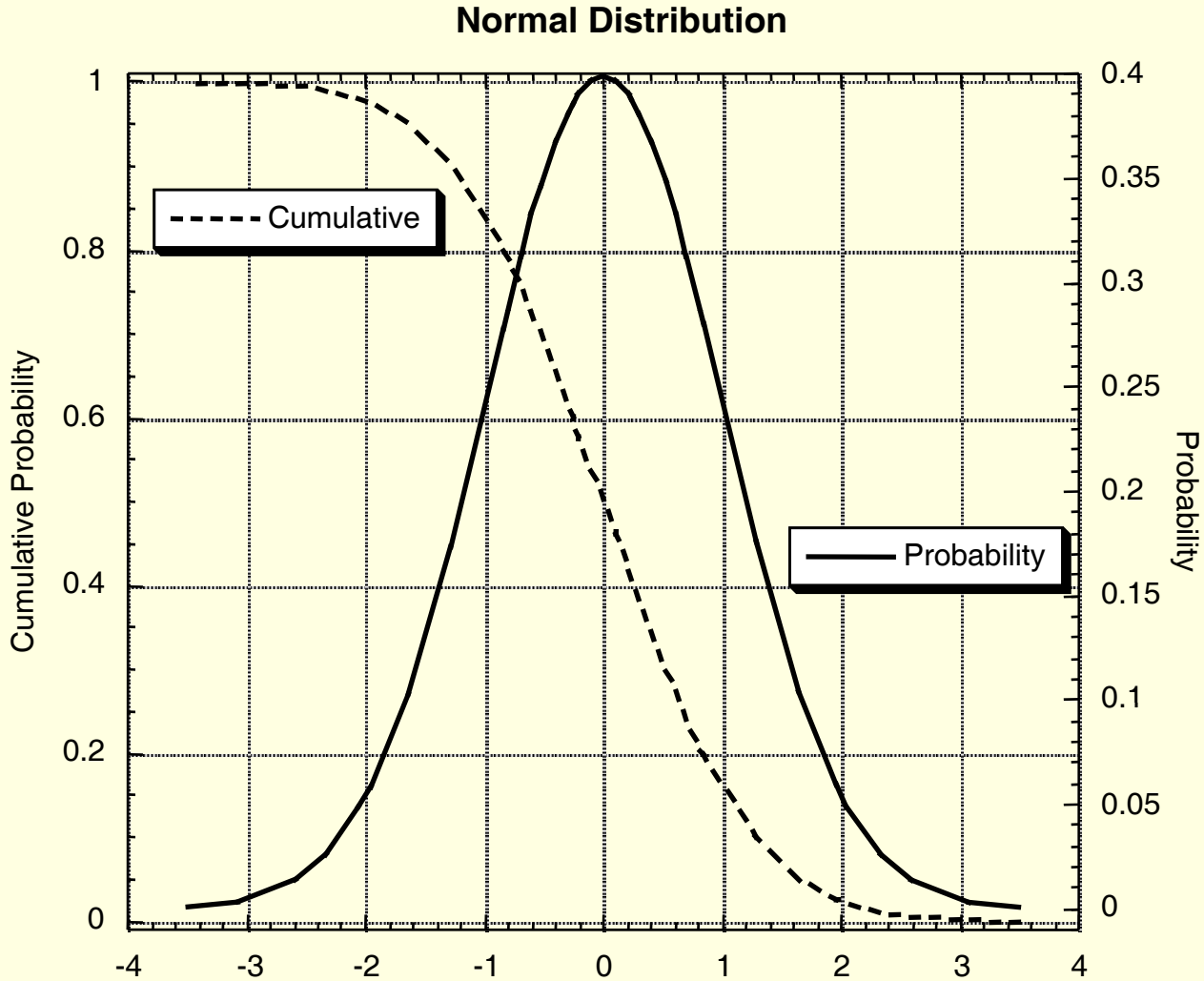
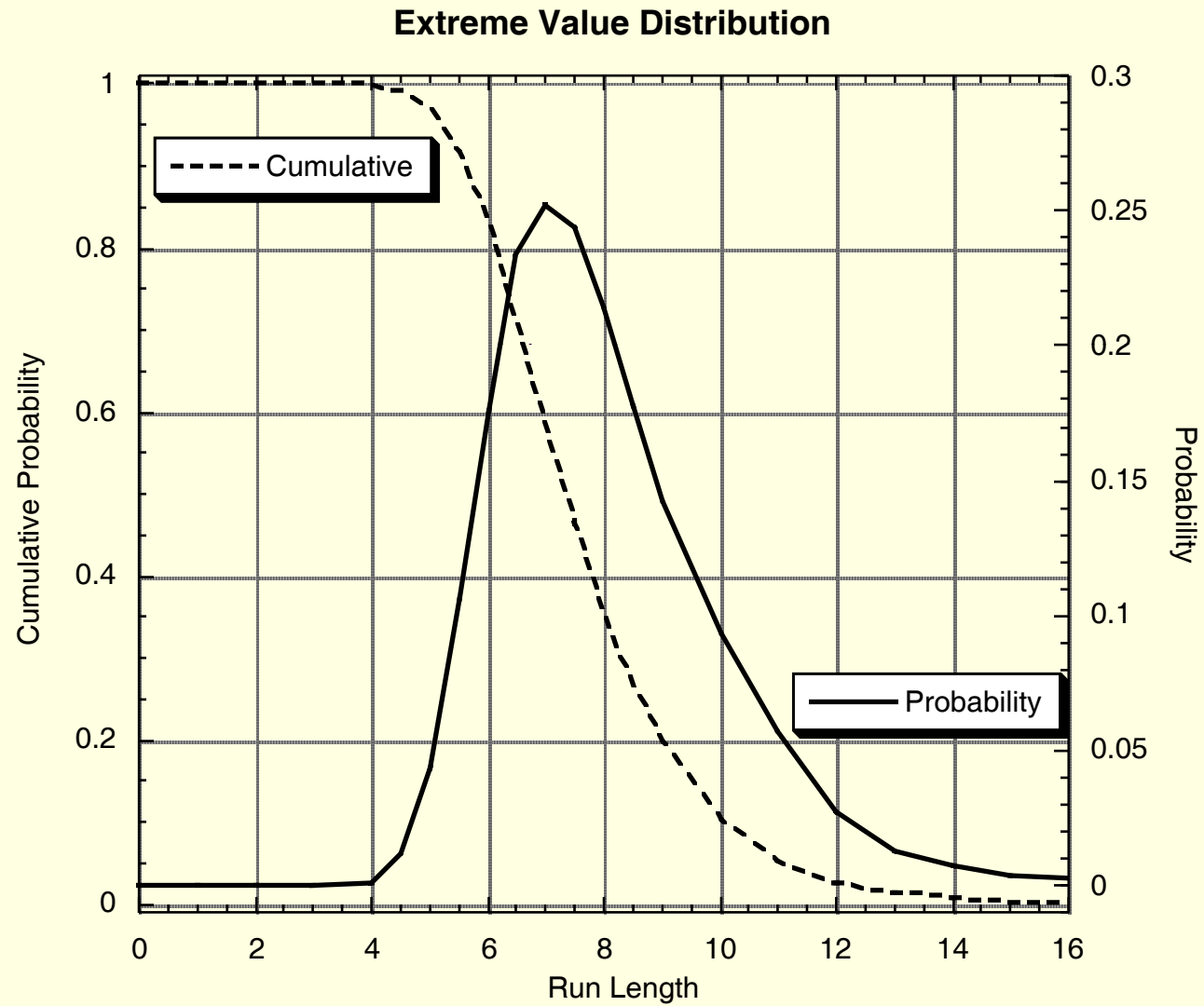


# *Preliminary Syllabus*

## Genomics

- Sep 30 Introduction & Genome Assembly
  - Oct 2 Sequence Comparison
  - Oct 7 Gene Modeling
  - Oct 9 Gene Function Identification – Read intro to HMM on blackboard
  - Oct 14 **OCTOBER BREAK**
  - Oct 16 Comparative Genomics
  - Oct 21 Protein-Protein Interactions
  - Oct 25 Pathway Resources and Analysis
  - Oct 28 Structural Genomics / Protein Structure Prediction
  - Nov 4 Protein Modeling
  - Nov 8 **EXAM**
- Gribskov@purdue.edu – Lilly G-233







### Goals

- Gene modeling begins with an uncharacterized genomic sequence and predicts the transcriptional and translational products of each gene, including
  - **Gene location, direction, and/or frame**
  - **5' and 3' untranslated regions**
  - **Introns and exons**
  - **Possibly includes regulatory elements**
- Gene modeling is notoriously difficult, especially in eukaryotes, but it is widely felt that current methods produce largely correct models, i.e. have errors in only 30% or so of eukaryotic genes and 10% of prokaryotic genes.
  - **Most common errors are in 5' end of gene and small exons**
  - **Difficult to distinguish errors from true genetic variation**
    - splice variants
    - pseudogenes

### *Basic Approaches*

- Prokaryotic genes are obviously easier
  - **No introns**
  - **Simpler signals**
  - **Often better DNA sequence**
- Eukaryotic genes are very challenging
  - **Exons/introns may be very small (less than 10 bases)**
  - **Introns may be very large (greater than 1 Mbase)**
  - **Signals are poorly known and more complex**
  - **DNA sequence may be more poorly assembled**

### *Basic Approaches*

- extrinsic – comparison to other known genes
  - **sequence comparisons to known proteins, cDNAs**
  - **genome comparison**
- intrinsic – properties of the sequence caused by the fact that it codes a protein
  - **ORF length**
  - **GC content**
  - **word frequencies**
- hybrid

### *Extrinsic methods (search by signal)*

- Try to identify sequence signals relevant to the presence, absence, frame, and content of genes
- Signals
  - **promoters**
  - **terminators**
  - **polyA sites**
  - **Cap signals**
  - **splice junctions**
- Sequence matches
  - **expressed genes (ESTs)**
  - **protein databases**
  - **closely related genomes (translated DNA vs translated DNA)**



### Sequence Motifs - Consensus Sequence

- Feature is represented as the majority or plurality character at each position

```
GCGGTGATAATGGTTGCATG
TTGGGTATATTTGACTATGG
ATGCATACACTATAGGTGTG
TGCAGTAAGATACAAATGGC
ATGGTTATAGTATGCCCATG
          TATAAT          GCGTG
```

### ***Sequence Motifs - Consensus Sequence***

- Advantages
  - **Concise**
  - **Simple to detect**
  - **Easily remembered and displayed**
- Disadvantages
  - **Most information is lost – poor ability to find signals**
  - **Difficult to evaluate partial match**
  - **Very sensitive to alignment**

### Sequence Motifs - Regular Expression

- Feature represented by logical combination of characters

GCGGT**G**A**T**AATGGTTGC**A**T**G**  
TTGGG**T**A**T**ATTTGACT **A**T**G**G  
ATGC**A**T**A**CACTATAGGT**G**T**G**  
TGCAG**T**AAGATA**C**AA **A**T**G**GC  
ATGGT**T**A**T**AGTATGCC**C**A**T****G**

          /  /  |  \  
          [ TG ] A [ TAC ] [ AG ] X T X ( 4 - 6 ) [ AG ] T G

### **Sequence Motifs - Regular Expression**

- Advantages
  - Fairly concise and easy to understand
  - Well known algorithms for matching,  $O(n \log n)$
  - Fairly easy to display
  - Can accept gaps
- Disadvantages
  - Still loses information, better than consensus
  - Rigid
  - Difficult to evaluate partial matches

### **Sequence Motifs - Regular Expression Methods**

- PROSITE Release 19.35, of 19-Sep-2006
  - **Constant updates**
  - **1331 different patterns, 4 rules and 650 profiles/matrices).**
  - **1446 documentation entries**
- Signatures derived by hand
- Relatively "fragile"
- Hulo N., Bairoch A., Bulliard V., Cerutti L., De Castro E., Langendijk-Genevaux P.S., Pagni M., Sigrist C.J.A.  
*The PROSITE database.*  
Nucleic Acids Res. 34:D227-D230 (2006)

### Sequence Motifs - Regular Expression Methods

- PROSITE “language”
- Each position is separated from the next by a hyphen “-”
- X means any residue
- [ ] surround ambiguities, e.g. [ALT] means ala, leu or thr
- { } surround forbidden residues, {AM} means neither ala nor met
- ( ) surround repeat counts
  - **(3)** means exactly three repeats
  - **(2-4)** means 2 to 4 repeats
- < and > indicate the beginning or end of the sequence, respectively
- . ends the pattern

### *Sequence Motifs - Regular Expression Methods*

- PROSITE – tabulated results are useful for training new methods
  - True positives (T) - Sequences that have the feature and match the signature
  - False positives (F) - Sequences that do not have the feature but match the signature
  - False Negatives (N) - Sequences that have the feature but do not match the signature
  - True negatives - Sequences that have the feature but do not match the signature
  - Potential (P) - likely to be a true positive
  - Maybe (?) - might have the feature, but unclear

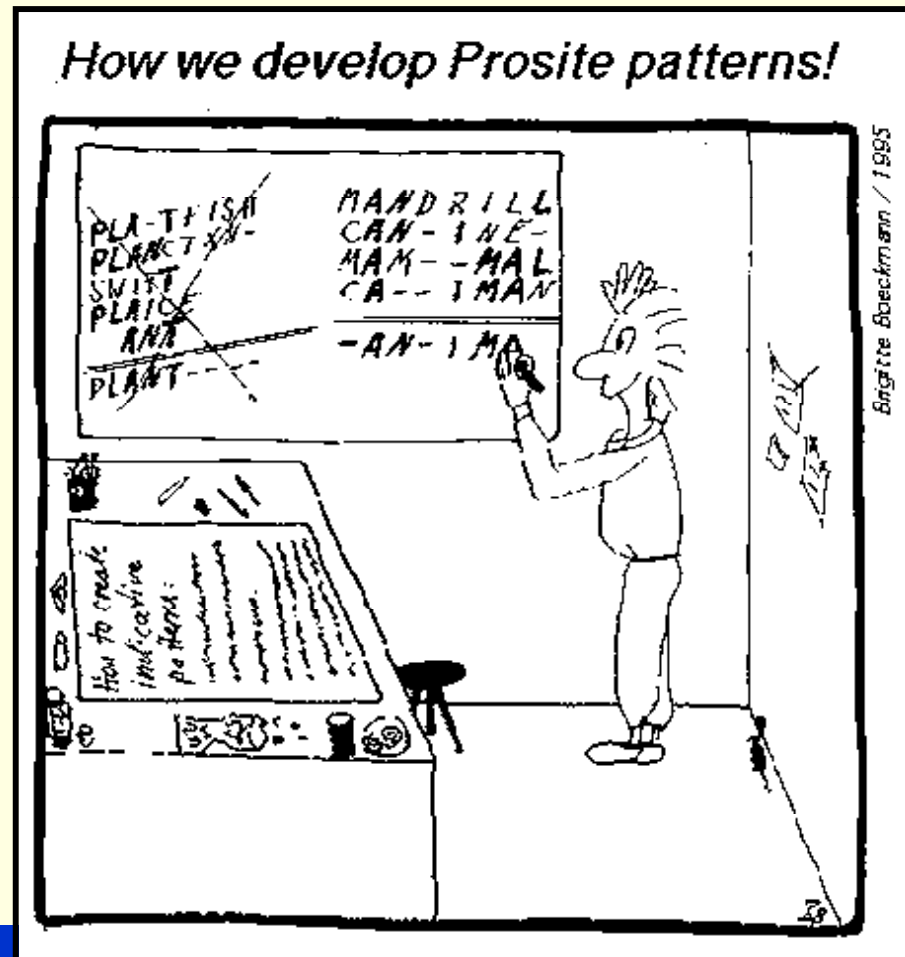
### Sequence Motifs - Regular Expression Methods

- A PROSITE entry

```
ID    CNMP_BINDING_2; PATTERN.
AC    PS00889;
DT    OCT-1993 (CREATED); OCT-1993 (DATA UPDATE); OCT-1993 (INFO UPDATE).
DE    Cyclic nucleotide-binding domain signature 2.
PA    [LIVMF]-G-E-x-[GAS]-[LIVM]-x(5,11)-R-[STAQ]-A-x-[LIVMA]-x-[STACV].
NR    /RELEASE=26,33329;
NR    /TOTAL=56(34); /POSITIVE=55(33); /UNKNOWN=0(0); /FALSE_POS=1(1);
NR    /FALSE_NEG=1(1);
CC    /TAXO-RANGE=??EP?; /MAX-REPEAT=2;
DR    P03020, CRP_ECOLI , T; P29281, CRP_HAEIN , T; P06170, CRP_SALTY , T;
DR    Q00194, CGCC_BOVIN, T; P29973, CGCC_HUMAN, T; P29974, CGCC_MOUSE, T;
DR    P05207, KAP2_PIG , N;
DR    P31324, KAP3_MOUSE, P;
DR    P29956, XANB_XANCP, F;
3D    2GAP; 3GAP; 1CGP;
DO    PDOC00691;
```



## Sequence Motifs - Regular Expression Methods



### *Sequence Motifs - Regular Expression Methods*

- PROSITE
- Steps to defining a signature (manual)
  - **1. Align sequences**
  - **2. Find a four or five residue sequence that is part of a known important region (core pattern)**  
Active site, substrate binding, prosthetic group, etc.
  - **3. Scan SWISS-PROT and see what matches**
  - **4. If only true positives are found, stop. Otherwise, add to the signature and return to step 3.**

### Sequence Motifs - Regular Expression Methods

- PROSITE
- Generation of signature - "Walker type" ATP binding sites

malk	SGCGKS.TLL
hisp	SGSGKS.TFL
oppd	SGSGKSQSRL
ecatpa	AGVGKT.VNM
bovatpb	AGVGKT.VFI

[SA]-G-[CSV]-G-K-[ST]-X(0,1)-[TSV]-[LMI]

- Simplest method - combine observed residues at each position

### Sequence Motifs - PSSM

- Position Specific Scoring Matrix, or weight matrix, is calculated based on observed frequencies in a column

```
GCGGTGATAATGGTTGCATG
TTGGGTATATTTGACTATGG
ATGCATACACTATAGGTGTG
TGCAGTAAGATACAAATGGC
ATGGTTATAGTATGCCCATG
```

### Sequence Motifs - PSSM

- Position specific scoring matrix (PSSM)
- Feature is represented as a matrix with a score for every possible character
- A simple weight matrix for the bacterial promoter -10 region, values here are simply % frequencies

<b>A</b>	<b>2</b>	<b>95</b>	<b>26</b>	<b>59</b>	<b>51</b>	<b>1</b>
<b>C</b>	<b>9</b>	<b>2</b>	<b>14</b>	<b>13</b>	<b>20</b>	<b>3</b>
<b>G</b>	<b>10</b>	<b>1</b>	<b>16</b>	<b>15</b>	<b>13</b>	<b>0</b>
<b>T</b>	<b>79</b>	<b>3</b>	<b>44</b>	<b>13</b>	<b>17</b>	<b>96</b>
	<b>T</b>	<b>A</b>	<b>T</b>	<b>A</b>	<b>A</b>	<b>T</b>

### **Sequence Motifs - PSSM**

- Advantages
  - Preserves first order information, i.e. assumes that positions are independent
  - Flexible, can model all regular expression type signatures
  - Accommodates partial matches, with known method for evaluating significance of matches
- Disadvantages
  - Difficult to display, impossible to remember

### Sequence Motifs - PSSM

- Log-odds matrix - as we have already learned, a log-odds statistic is one of the most powerful discriminators. Weight matrices are often in log-odds form.

$$w_{ij} = \ln ( f_{obs} / f_{exp} )$$

$$score = \sum w \text{ over width of pattern}$$

- What should one use for the background model,  $f_{exp}$  ?
  - Database composition
  - Global composition of query sequence
  - Local composition of query sequence
  - Combination of query and database sequences

## Search by site

- Eukaryotic transcription initiation site

	-3	-2	-1	0	1	2	3	4	5	6	7	8	9	10	11
A	16	4	90	1	91	69	92	57	40	14	21	21	21	17	20
C	37	12	0	2	0	0	1	1	11	35	38	33	30	28	26
G	39	5	1	1	1	0	5	11	40	39	33	33	33	36	36
T	8	79	9	96	8	31	2	31	9	12	8	13	16	19	18

-----

G	T	A	T	A	A	A	A	G	G	C	G	G	G	G
S	T	A	T	A	W	A	W	R	S	S	N	N	S	S

%frequency per position

Y = pyrimidine = C or T  
 R = purine = A or G  
 S = strong = G or C  
 W = weak = A or T



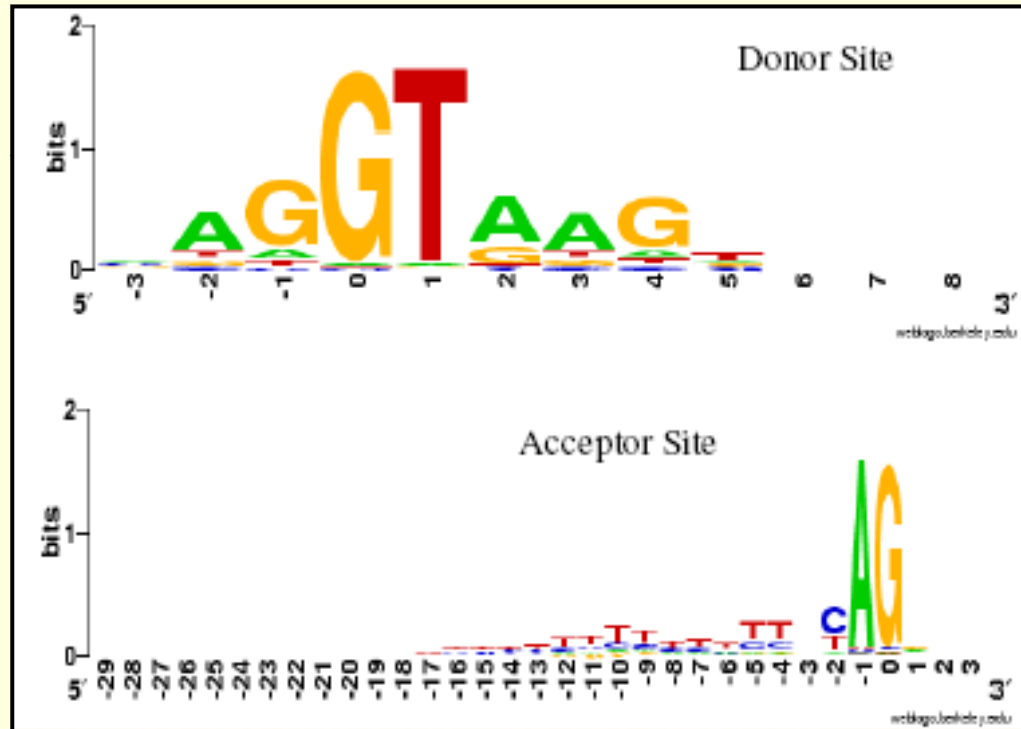
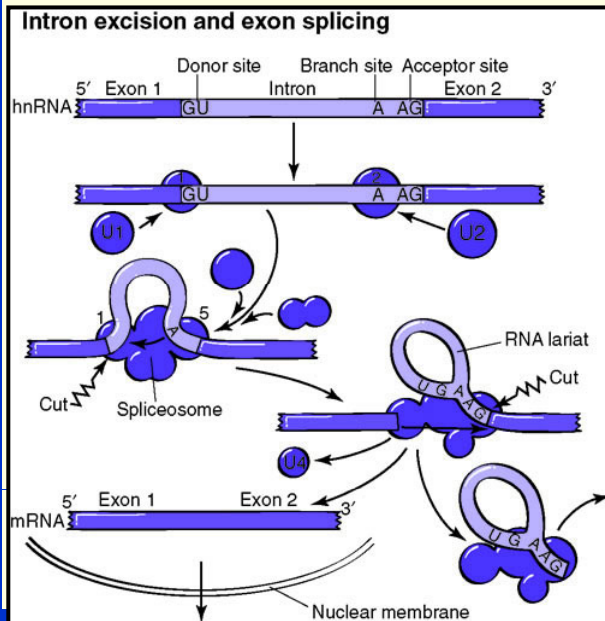
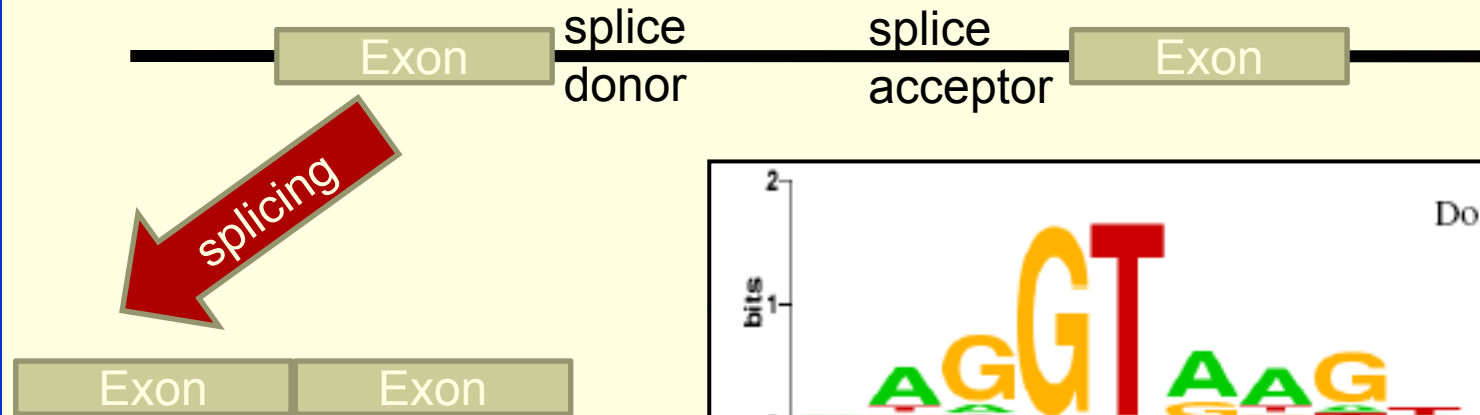
### Search by Site - Splice sites

- The splicing of introns is a multi step process of RNA maturation which takes place in the nucleus
  - **generate mature mRNA molecules for transport to the cytoplasm.**
  - **Involves a complex of several factors such as snRNP (small nuclear ribonucleoprotein particles) and hnRNPs (heterogeneous nuclear ribonucleoprotein particles). This complex assembly is called the spliceosome.**
- Introns usually begin with GU (donor splice site) and end with AG dinucleotides (acceptor splice site).
- The branch point signal typically is located 10-50 bases upstream from the acceptor splice site (the lariat region).

# Genomics - Gene Modeling

Genomics

## Splice signals



mouse splice junction

# Genomics - Gene Modeling

## Search by Site – Splice junction

### • Donor site

A	28	59	8		0	0	54	74	5	16
C	40	14	5		0	0	2	8	6	18
G	17	13	81		100	0	42	11	85	21
T	14	14	6		0	100	2	8	4	45
	C	A	G		G	T	A	A	G	T

### • Acceptor site

A	10	8	6	6	9	9	8	9	6	6	23	2	100	0		28
C	31	36	34	34	37	38	44	41	44	40	28	79	0	0		14
G	14	14	12	8	9	10	9	8	6	6	26	1	0	100		47
T	44	43	48	52	45	44	40	41	45	48	23	18	0	0		11
	T	T	T	T	T	T	T	T	T	T	N	C	A	G		G



Position Specific Scoring Matrix (PSSM)  
or Weight Matrix

### Search by Site – splice signals

- Branch point signal

A	1	0	39	99	11
C	76	8	15	1	45
G	2	0	42	0	6
T	21	91	4	0	38
	C	T	G	A	C

A	-5.8	-6.8	-0.5	0.8	-2.3
C	0.8	-2.5	-1.3	-5.5	0
G	-4.5	-6.5	-0.1	-6.5	-2.9
T	-1.4	0.7	-3.8	-6.8	-0.5
	C	T	G	A	C

Consensus: CTGAC

Regular Expression: [CT]T[AG]A[CT]  
YTRAY

Y = pyrimidine = C or T

R = purine = A or G

S = strong = G or C

W = weak = A or T

Log-odds assuming 45% AT, 55% GC

### Search by Site

- Eukaryotic translation initiation site

	-6	-5	-4	-3	-2	-1	+1	+2	+3
A	18	19	24	68	23	15	100	0	0
C	21	40	58	2	55	53	0	0	0
G	47	23	12	30	16	23	0	0	100
T	13	18	6	0	7	9	0	100	0
	G	C	C	A	C	C	A	T	G

### Search by Site

- Consensus sequences

- **CCAAT-box**

- Y Y Y R R C C A W W S R -212 .. -57

- **GC-box**

- W R K R G G Y R K R K Y Y K -164 .. +1

- **cap-site**

- K C W K Y Y Y Y +1 .. +5

- **Information about composite regulatory elements, transcription factors and eukaryotic promoters are collected in the following databases:**

- TRANSFAC, <http://www.gene-regulation.com/pub/databases.html> (Wingender et al., 1996).

- TFD, <http://www.ifti.org/ootfd/>  
(Ghosh, 1993)

- EPD, epd promoter, (Bucher, 1988)

### *Search by Site*

- Polyadenylation site
- Polyadenylation (cleavage of pre-mRNA 3' end and synthesis of poly-(A) tract) is a very important early step of pre-mRNA processing.
- Sites
  - **AATAAA**, located **15-20 nucleotides upstream from the poly-(A)**
  - **ATTAAA**, is nearly as active as the canonical sequence.
  - An additional signal with consensus **YGTGTTY** (diffusive GT-rich sequence) was revealed in region from 20 to 30 nucleotides downstream of poly-(A) site (site of cleavage) (McLauchlan et al., 1985).

### *Search by Sites*

- Methods for identifying sites (weakest to strongest)
  - **Consensus sequence**
  - **Regular expression**
  - **Log-odds matrix / window analysis (PSSM)**
  - **Neural network or Hidden Markov model**



## *What is Homology?*

- Nothing in biology makes sense except in the light of evolution.
  - **Theodosius Dobzhansky (1900-1975)**
  - **...without that light it becomes a pile of sundry facts some of them interesting or curious but making no meaningful picture as a whole.**
- **homology** - the presence of a similar feature because of descent from a common ancestor
- **homoplasy** - the presence of a similar feature because of convergence
  - **Homology cannot be observed. We can't actually see the ancestral organisms/molecules and trace descent.**
  - **Homology is an inference, a conclusion we draw based on observed similarity.**
  - **Homology is an all-or-none relationship**

### *Why is homology Important?*

- Homology strongly suggests that the molecules have similar structure and function
- There are (very) many ways to fold a polypeptide to place specific chemical groups at specific locations. There is no reason, *a priori*, why proteins with a specific function should have similar 3-D structures.
- Therefore, there is no reason, *a priori*, why unrelated sequences should have any detectable similarity in sequence. Significantly similar molecular sequences are very unlikely to arise by chance - i.e. homoplasy on the molecular level is very unlikely.
- When we see significant similarity, we infer that the sequences/structures are homologous, i.e. at some point in the past they share an identical structure.
- The only thing that keeps the sequences tied to each other is the commonality of structure and function arising from homology.

### Homology

- Sequences alignments and database searches let us
  - **Find homologous sequences (genes/proteins)**
  - **Map information from known systems to new ones**
    - Gene identification
    - Gene function
    - Metabolic and regulatory systems
- Two common classes of homologs
  - **Orthologs – genes separated by a speciation event, i.e. the same gene in two species**
  - **Paralogs – genes separated by a duplication events, originally the same but now diverged with possibly different functions**

### *BLAST Basic Idea*

- Determine in advance the MSP score you need to be significant,  $S$ 
  - for example, choose  $S$  so that you will see fewer than 10 unrelated sequences in the database that score as high
- Look for matching words of length  $w$  that score above a threshold,  $T$ , such that MSPs of score  $S$  are unlikely to be missed. These are High-scoring Segment Pairs (HSPs)

### *BLAST procedure*

- Step 1: Compile list of high scoring words from query
- Step 2: Scan database for "hits"
- Step 3: Extend regions with 2 hits into MSPs
- Step 4: Dynamic programming alignment around MSPs

sequence

### *BLAST Step 1 - List of High Scoring Words*

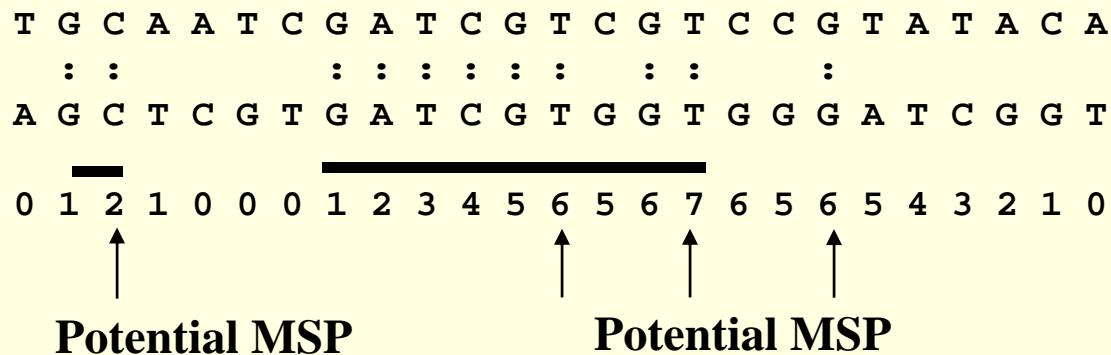
- Choose a significance level  $S$
- Choose a word size,  $w$ , and cutoff,  $T$ , so that you are unlikely to miss MSPs with score  $S$
- Make a table of all words in the "neighborhood" of the query (DNA sequences use all words)
- Typically 50 words for each residue

### *BLAST Step 2 - Scan Database*

- Scan only for words in neighborhood
- Use lookup tables (like FASTA) or finite automaton
- Keep data in memory to make it faster

### BLAST Step 3 - Extend Words to MSPs

- In BLAST2, a “diagonal” must have two word hits before extension to MSP is attempted.
- In principal, must examine diagonal until score drops to zero
- Shortcut, only check until score drops by X





### *Filtering*

- Some sequences give spurious matches because of their unusual properties. Such sequences are automatically filtered by BLAST
- Filters remove “low entropy” sequences. These are repetitive sequences that often give anomalous matches in a database search.
  - **Degenerate sequences – e.g., poly A runs**
  - **Dinucleotide, trinucleotide (or longer) repeats**
  - **Transmembrane regions and signal peptides in proteins**